

Research Article

GASTROINTESTINAL HELMINTH INFECTION OF LABORATORY MICE AND WILD RODENTS IN AIZAWL, MIZORAM

GHOSH SUBHAMOY¹, PATRA GAUTAM^{1*}, ROY LEENA², BORTHAKUR SONJOY KUMAR¹ AND LALRINKIMA H.¹

¹Department of Veterinary Parasitology, College of Veterinary Sciences and Animal Husbandry, Selesih, Aizawl, India ²Department of Veterinary Public Health, College of Veterinary Sciences and Animal Husbandry, Selesih, Aizawl, India *Corresponding Author: Email- gautampatra488@gmail.com , sanjoy_barthakur@rediffmail.com

Received: October 31, 2016; Revised: December 24, 2016; Accepted: December 25, 2016; Published: December 28, 2016

Abstract- The aim of this study was to investigate various helminth infections of laboratory mice, musk rats as well as in wild rats which were trapped from various areas of Aizawl. The study showed that the laboratory mice were infected with mostly two species of cestodes and one species of nematode. The musk rats were found to be infected with one species of cestode namely *Rodenolepis microstoma* and one species of nematode, *Capillaria hepatica*. However, *Trichinella sp* larvae could be recovered from muscle of wild rats. The prevalence of helminthic infection in laboratory mice were *Hymenolepis diminuta* (30%), *Hymenolepis nana*,(20%), *Cysticercus fasciolaris* (5%) and *Syphaciamuris* (10%), respectively. In musk rats, *Rodenolepes microstoma* (5%), *Capillaria hepatica*(5%) andin wild rats *Trichinellas*plarvae(1%) were recorded. Since most of the helminths found in rodents are of zoonotic importance, the results suggest that rats either domestic or wild may be acted as a source of helminth transmission to human in this region of India.

Keywords- Rodents, Helminths, Zoonosis, Aizawl, Mizoram.

Citation: Ghosh Subhamoy, et al., (2016) Gastrointestinal Helminth Infection of Laboratory Mice and Wild Rodents in Aizawl, Mizoram. International Journal of Parasitology Research, ISSN: 0975-3702 & E-ISSN: 0975-9182, Volume 8, Issue 5, pp.-194-196.

Copyright: Copyright©2016 Ghosh Subhamoy, et al., This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution and reproduction in any medium, provided the original author and source are credited.

Academic Editor / Reviewer: Suman Kundu

Introduction

Rats besides causing great damage to the ecosystem can also harbour a variety of helminths of zoonotic importance. Some previous studies have been done on the occurrence of helminths in rodents in India [1-3]. However, little is known about parasite diversity in laboratory mice and wild rodents in Mizoram where wild rat populations are very high. It is well recognized that they are the hosts of various helminths and because of close proximity to human they pose a great threat to health hazard to human being [4]. The dissemination of parasitic infection takes place when faeces containing eggs are passed along with the rodent droppings and contaminating agricultural lands, stored grains, water sources and in various edible things [5]. In Mizoram, 31% of the total forest area is covered by bamboo forests and the outbreak of rodents population is a common phenomenon particularly during bamboo flowering. There are several reports on the occurrence of the helminths in rodents across the world [6-8] which clearly indicate the wide diversity of helminths in these animals.

The present study is therefore underscores helminth biodiversity among rodents in correlation with habitat so that an evaluation of the risk for helminth transmission to humans can be ascertained. We examined laboratory reared mice as well as trapped musk rats and wild rats in different locations of Aizawl district, Mizoram.

Materials and Methods:

A total number of 100 laboratory mice and 20 numbers of musk rats were examined. We were also able to capture 20 wild rats altogether through trapping device. The musk rats were captured near bamboo forests, rice fields, house yards, godowns and from animalbarns. Cotton wool soaked in chloroform was used for induction of anaesthesia followed by euthanasia of the musk rats and wild rats. Five dead laboratory mice were also brought to the laboratory for post-mortem examination. Each visceral organ was looked carefully for recovery of

helminth parasites. The parasites recovered were cleared in 0.85% physiological saline with the help of camel brush several times before undergoing whole mount permanent preparation as per standard procedures. Cestodes were transferred from 70% alcohol and stained in Semichon's carmine stain for 2-4 hours. Then the specimens were transferred to 70% alcohol for 20-30 minutes. The parasites were then destained in 70% acid ethanol for 30 seconds to 15 minutes depending upon the stain, the size and type of the worm . In the next step, the specimens were put into 70% basic ethanol for 30 seconds to 15 minutes, transferred to 70% ethanol for 10 minutes and then 95% ethanol for 20-30 minutes and then in 100% ethanol with 2-3 changes for 20-30 minutes each. Then the specimens were cleaned in xylene with two changes for 20-30 minutes each. Finally, the specimens were permanently mounted on Canada Balsam. The nematodes were isolated and preserved in 70% alcohol. Nematodes were cleaned with lactophenol and mounted on a temporary slide. The prevalence of particular parasite in a particular rat and parasite intensity in each infected rat were recorded. Identification of the parasites were do ne using standard keys. Faecal samples were also examined by conventional method for the detection of helminth eggs and identifies as per Souls by (1982) [9]. The muscle samples of 20 wild rats were digested into pepsin solution after thorough mincing with fine scissors.

Results:

The results of the distribution of parasites in different hosts and their locations. The different parasite recovered and the eggs of helminths and [Fig-1(a-c)]. Out of 100 numbers of laboratory mice examined, 10(10%) were found to be positive for *Syphacia muris* [Fig-2], 30(30%) were positive for *Hymenolepis diminuta* [Fig-3], 20 (20%) were positive for *Hymenolepis nana* [Fig-4] and 5(5%) were positive for *Cysticercus*

fasciolaris [Fig-5]. From 20 muscle samples, one was positive for *Trichinella sp* larvae [Fig-6]. Out of 20 musk rats examined, 2 (10%) were found positive for *Rodenolepes microstoma* [Fig-7]. One musk rat showed a large number of *Capillaria hepatica* in liver and pancreas [Fig-8]. No trematodes or acanthocephalan could be recovered in the present investigation.



Fig-1a Hymenolepis diminuta ova



Fig-1b Syphacia muris ova



Fig-1c Hymenolepis nana ova



Fig-1d Capillaria hepatica egg



Fig-2 Syphacia muris



Fig-3 Scolex of Hymenolepis diminuta



Fig-4 Hymenolepis nana



Fig-5 Cysticercus fasciolaris



Fig-6 Rodenoleptis microstoma

International Journal of Parasitology Research ISSN: 0975-3702&E-ISSN: 0975-9182, Volume 8, Issue 5, 2016



Fig-7 Trichinella sp



Fig-8 Capillaria hepatica in liver of musk rat

Discussion

Rodents have been studied for gastrointestinal helminths in different parts of India as well as different parts of world but scanty information regarding the load of helminths in rodents are available from North-Eastern region of India. In present study, the predominant species of cestode was Hymenolepis diminuta followed by Hymenolepis nana and Cysticercus fasciolaris in the liver of laboratory mice. Among nematodes, only 10 mice were found positive for Syphacia muris. Malsawant luangi and Tandon also recorded almost similar percentage (9.37%) of Syphacia muris from Rattus nitidus from this region [3]. The high prevalence of Hymenolepis diminuta in mice agreed to that recorded by other co-workers [1,10]. One species of cestode have been recovered from wild musk rats namely Rodenolepes microstoma. This is the first time to the best of our knowledge that Rodenolepis microstoma has been found in the GI tract of wild rodents in India. None of the previous studies by various Indian workers have reported Rodenolepis microstoma in the GI tract of rodents. However, molecular identification required for validity of this species. Only one musk rat was found positive for Capillaria hepatica infection at post-mortem examination. However, rats were least commonly infected with Trichurismuris and Capillaria spp [2]. One muscle sample was positive for Trichenella splarvae, which require molecular technique to identify the actual species involved.

The absence of any trematode in the present investigation is clearly understood in the sense that trematodes need aquatic snails for the completion of their life cycle and terrestrial habitat of rodents in this part of country preclude the possibilities of such infection. The study provides basic information on GI helminths of rodents from one of the North-Eastern parts of India. However, further studies are needed in other parts of North-Eastern states in order to analyse the potential of helminth zoonos is in India.

Conclusion

Rat and mice harbour several helminth parasites which not only affect it's host but also pose for threat to human health.

Conflict of interest statement

We declare that we have no conflict of interest.

Author Contributions

SG and LR prepared the initial version of the manuscript and conducted

Laboratory works. SKB and LK helped in the collection of literature and supervised the work and GP donethe scientific and technical corrections.

Acknowledgements

The authors duly acknowledged the Dean, College of Veterinary Sciences & A.H., Central Agricultural University, Selesih, Aizawl, Mizoram for providing necessary facilities to conduct the study.

Abbreviations

SG: Subhamoy Ghosh LR: Leena Roy SKB: Sonjoy Kumar Borthakur GP: Gautam Patra LK: Lalrinkima

Ethical approval

This article does not contain any studies with human participants or animals performed by any of the authors.

References

- [1] Sharma D., Joshi S., Vatsya S. and Yadav C.L. (2012) J Parasit Dis.
- [2] Singla L.D., Singla N., Parshad V.R., Juyal P.D. and Sood N.K. (2008) Integr Zool, 3, 21-26.
- [3] Malsawmtluangi C. and Tandon V. (2009) J Parasit Dis, 33(1-2), 28-35.
- [4] Zain S.N.M., Behnke J.M. and Lewis J.W. (2012) Parasites Vectors, 5, 47.
- [5] Khatoon N., Bilqees F.M., Shahwar D. and Rizwana A.G. (2004) *Tur J Zool*, 28, 345-351.
- [6] Claveria F.G., Causapin J., de Guzman M.A., Toledo M.G. and Salibay C. (2005) Southeast Asian J Trop Med Public Health, 36(4).
- [7] Gomez Villafane I.E., Robles M.R. and Busch M. (2008) *Helminthologia*, 45(3), 126-129.
- [8] Chaisiri K., Chaeychomsri W., Siruntawineti J., Ribas A., Herbreteau V. and Morand S. (2010) J Trop Med Parasitol, 33, 29-35.
- [9] Soulsby E.J.L. (1982) Helminths, Arthropods and Protozoa of Domestic Animals, 7th edition.
- [10] Gudissa T., Mazangia H., Alemu S. and Nigussie H. (2011) J Infect Dis Immun, 3, 1-5.