

Organic constituents in the different stages of regenerated callus of sugarcane Var Co-740

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Abstract- In all living organisms there is an increasing procession of chemical reactions and processes, which leads to fundamental internal changes. Living substance is constantly being torn down and built up. The general activities of protoplasm are being maintained. Waste products are being eliminated and vast amount of energy is constantly being expended. The sum total of these processes and changes is called metabolism. The present investigation was undertaken to study the changes in organic metabolites of different regenerated stages in sugarcane var. co 740. The organic matters like chlorophyll, protein, proline, and polyphenols, reducing sugar, total sugar starch, moisture percentage and TAN were analyzed.

Keywords- Sugarcane, metabolites, organogenesis, proline

Introduction

Sugarcane is one of the cash crops of India as 60% of the world sugar. Sugarcane industries is the second largest agro based industry [7]. Sugarcane (*saccharum* species hybrid) tissue is widely used in sugarcane improvement and breeding programs [2, 5, and 8]. Callus can be initiated from any sugarcane tissue [6]. Organogenesis is the development of adventitious organs from undifferentiated cell mass in tissue culture by the process of differentiation [10, 12]. It has found that addition of auxins to the culture medium to stimulate root formation. Skoog and Miller have reported that organogenesis is controlled by a balance between cytokinin and auxin. Organogenesis studies in vitro conditions helps in understanding the developmental biology of sugarcane tissue culture at metabolic and biochemical level.

Materials and methods

Three different stages of tissue culture plant material were used for the investigation.

Estimation of Chlorophylls

Total chlorophylls were estimated by the method of Arnon [1] in which 1 gm of fresh plant material of three different stages was crushed in cold mortar using pestle in 80 % (v/v) acetone mixed with 2 ml of ammonia (25%) per liter. During extraction, a pinch of acid washed sand 0.10 gm of MgCo₃ was added. Extraction was carried in the dark and cold condition. The extract was filtered through Buchner funnel and used for spectrophotometric determination of absorbance at 663 and 645 nm. Total chlorophylls were calculated by standard formula.

Chlorophylls a+b= $8.02 \times \text{Absorption at } 663 + 20.2 \times \text{Absorption at } 645 [Z]$

Total chlorophylls= $\frac{Z \times \text{VOLUME OF EXTRACT} \times 100}{\text{WEIGHT OF PLANT MATERIAL} \times 100}$

Estimation of Polyphenols

Polyphenols are estimated from the various parts using the oven dried powdered material by Folin Denis method [1, 4]. Polyphenols from dried plant material were extracted in 80% acetone-30ml. The extract was filtered through Whatman no.1 filter paper using Buchner funnel under suction. Polyphenols were extracted repeatedly from the residue. The volume of the filtrate was made to 50 ml. The same filtrate was used for estimation of polyphenols. 2 ml of filtrate was taken in 50 ml marked Nessler's tube. In other such tubes different concentration (0.5, 1.0, 2.0 and 4.0 ml) of standard polyphenol solution (tannic acids 0.1 mg/ml) were taken 10 ml of 2% Na₂CO₃ were then added to each test tube to make the medium alkaline and 2 ml of Folin Denis reagent.

A blank was prepared without polyphenols solution. The ingredients were allowed to mix thoroughly and after 10 min the optical density of each mixture was read at 660nm on spectrophotometer.

Estimation of Proline

The estimation was carried out by the method of Bates [11]. About 0.5g oven dried plant material was homogenized in 10ml sulphosalicylic acid (3%) and the extract was filtered through whatman no.1 filter paper. The assay known quantity of filtrate was mixed with 2 ml of acid ninhydrin reagent. The contents were boiled for 1 hour on boiling water bath and cooled rapidly in freeze ice bath. 4 ml of toluene was added to each test tube and vigorously shaken for few seconds. The absorbent of toluene chromophere was recorded at 520nm against toluene as blank. Standard curve of proline was prepared for different concentrations for proline estimation.

Estimation of Nitrogen and Protein

The total nitrogen was estimated by the method of Hawk [1, 3]. About 1 gm of oven dried plant material was taken in kjeldha's flask and 10ml of 1:1 H₂SO₄ and pinch of micro salt(CuSO₄+K₂S₄) 1:5 was added. The flasks were kept overnight. Next day the material was digested on hot plate until the fumes subside and a colorless acid digested extract was obtained. The extract was used for estimation of nitrogen colorimetrically at 415nm using Nessler's reagent. Total protein was estimated by Bradford method (1976). About 1 gm of fresh sample was homogenized in 0.1N NaOH and centrifuged at 2000 rpm, for 15 mins. The supernatant was used for determination of total protein. The absorbance read at 595 nm with bovine serum albumin was used as standard.

Estimation of Total Soluble Sugars

The total sugar was estimated by Anthrone method [1, 4]. About 1 gm of material was homogenized in 80% alcohol. The homogenate was centrifuged at 2000 rpm for 15 mins. The alcohol was evaporated on a hot water bath and filtrate was used. The final volume was adjusted to 10ml by adding distilled water and the extract was used for analysis.

Estimation of Sugars

Reducing sugars were estimated by Somogyis method [1, 9]. The alcohol evaporated extract was used for analysis.

Estimation of Titrable Acid Number (TAN)

Titration acid number was determined by modified method of Thomas and Beevers [6]. About 1 g of callus from different stages were boiled in 100ml of distilled water for 1 hr. and the filtered through muslin cloth. The filtrate was used for titration. To 10ml of extract 25ml of distilled water and 2-3 drops of phenolphthalein indicator was added in an evaporating dish. This was titrated against N/40 NaOH. The TAN is expressed as the ml of decinormal alkali required to neutralize the acids present in 100g of material.

Results

Chlorophyll Contents

The results of total chlorophyll contents in different stages of regenerating callus are presented in Table 1 and Figure 2. The callus at the initial stage showed only 10.3 mg of chlorophyll. As regeneration advances due to formation shoot buds the chlorophylls contents has shown linear increase. In the medium green stage, the amount has doubled within the 3-4 weeks of advancement of organogenesis process. This is about 267% more as compared to medium regenerated stage (IM). Thus our results show that chlorophyll synthesis is rapid in the later stages of regeneration response than at the initial stage.

Polyphenols

The Table-2 and Figure-4 shows the contents of polyphenols in three stages of callus cells. The polyphenols contents shoe gradual decrease as the cells become green and then develop into shoots. The callus had 912mg of polyphenols which declined to 460 and 210mg/100g as they turn medium green and into green shoot buds respectively.

Proline

The quantity of proline contents is shown in the same Table -2 and Figure-3. It is one of the important amino acid during growth and development during stress conditions. The proline contents are reduced in the medium green cells while it increases cells in the completely regenerating green seedlings.

Sugars

Sugars are important constituents of the growing cells. Reducing sugar level in the medium green cells is higher by 55% than in the non regenerated callus while it is reduced by 37% in the completely green callus. Similar trend is observed with respect to total sugar and starch contents. It seems that the sugar accumulates in the initially regenerating phase and it is rapidly utilized in the later process of organogenesis. This effect is more pronounced in starch contents of the cell as compared to total soluble sugars as shown in Table-3 and Figure -5.

Nitrogen and Protein

Organic constituents from three different stages of regenerating callus of sugarcane were analyzed. The results are shown in Table-2 and Figure 3 and 4. The total nitrogen content showed linear increased by 63.5% in medium green than in the non-regenerating callus by 29% in the complete regenerating seedlings. Similar trend is also seen in protein contents as shown in figure-4. As the cells undergo organogenesis, more nitrogen contents are assimilated and used for protein synthesis.

Total Acidification Number

TAN is defined as a number of decinormal alkali required to neutralize acids present in 100g of fresh tissue. It is can be seen in Table 1 and Figure 2. The acidity values are similar in callus and regenerated shoots. However, TAN has shown a higher level (16%) in the medium green callus. These results again indicate that the metabolic status of medium green tissue is more active as compared to the non-regenerated cell and regenerated green shoots. The medium green cells along with acquiring high water content are also maintained higher acidity. It seems that the cells from this stage are active in generating more ATP through Krebs cycle and hence show elevated acid level.

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Table 1- Moisture, TAN, Chlorophyll contents in the different stages of regenerated shoots of sugarcane variety Co 740

Stages	Moisture (%)	Total chlorophyll (mg/100g)	TAN
IC	92.0(±6.8)	10.3(±2.8)	70.6(±8.7)
IM	93.6(±5.2)	24.4(±5.4)	82.3(±6.5)
IR	95.0(±5.6)	89.6(±10.6)	72.4(±3.5)

Table 2-Organic constituents in the different stages of regenerated shoots in sugarcane variety Co 740

Stages	NITROGEN ((mg/100mg)	PROTEIN (mg/100mg)	POLYPHENOLS (mg/100mg)	PROLINE (mg/100mg)
IC	0.52(±0.12)	303.2(±2.7)	912(±68.2)	6.6(±1.2)
IM	0.85(± 0.17)	492.0(±28.3)	460(± 72.5)	5.2(±1.7)
IR	1.10(±0.28)	644.8(±25.2)	210(±27.3)	8.2(±0.75)

Table 3- Carbohydrates in the different stages of regenerated shoots of sugarcane variety Co 740

STAGES	REDUCING SUGAR (mg/100g)	TOTAL SUGAR (mg/100g)	STARCH (mg/100g)
IC	5.52	6.04	7.76
IM	8.55	12.83	2.8
IR	5.45	8.68	3.4

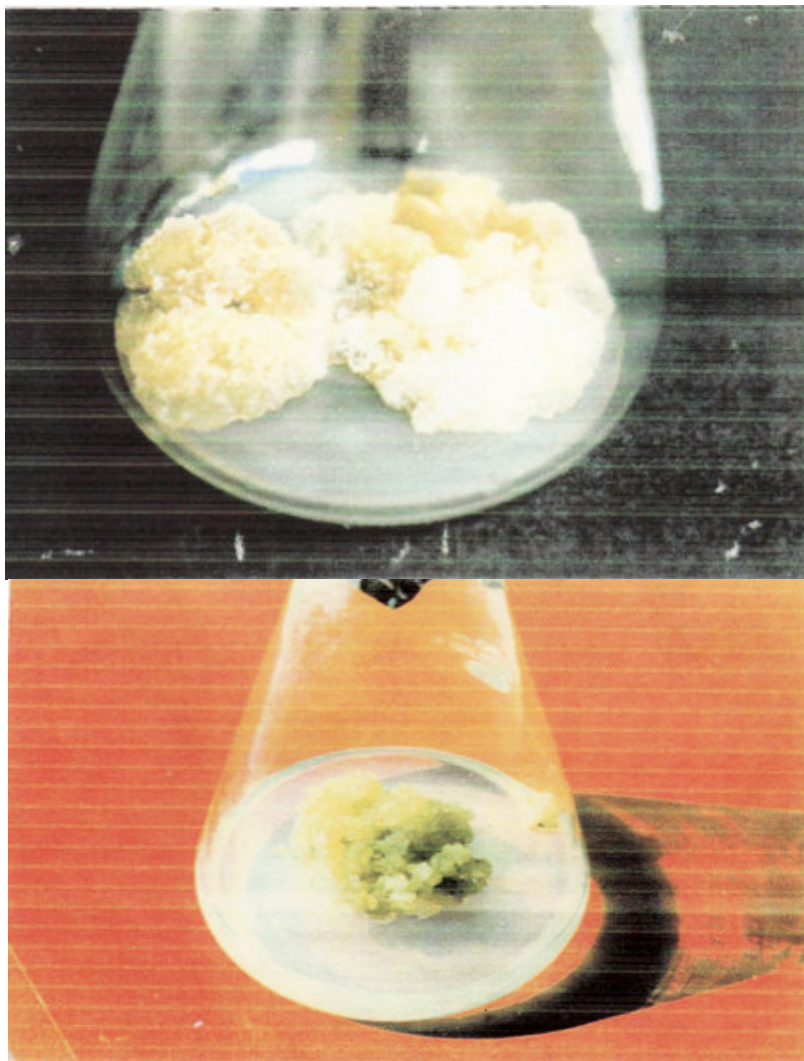


Fig.1- Regenerated callus in sugarcane Variety Co-740

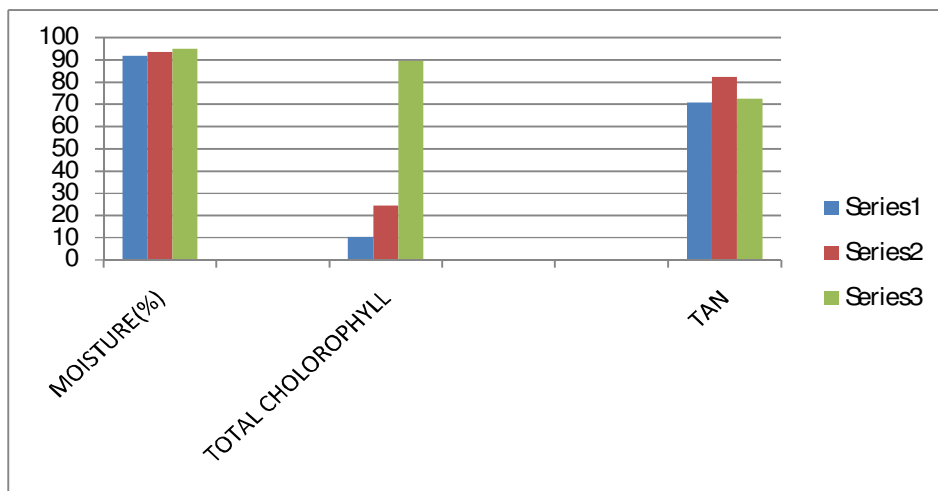


Fig. 2- Organic constituents in the different stages of regenerated shoots in sugarcane variety Co.740 (Series 1: IC, Series 2: IM and Series 3:- IR)

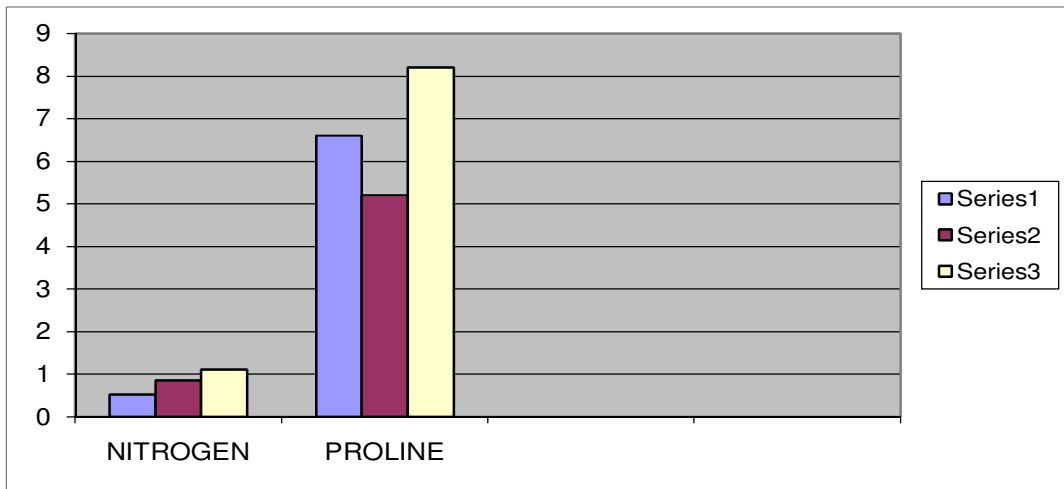


Fig. 3- Organic constituents (nitrogen and Proline) in the different stages of regenerated shoots in sugarcane variety Co.740 (Series 1: IC, Series 2: IM and Series 3:- IR)

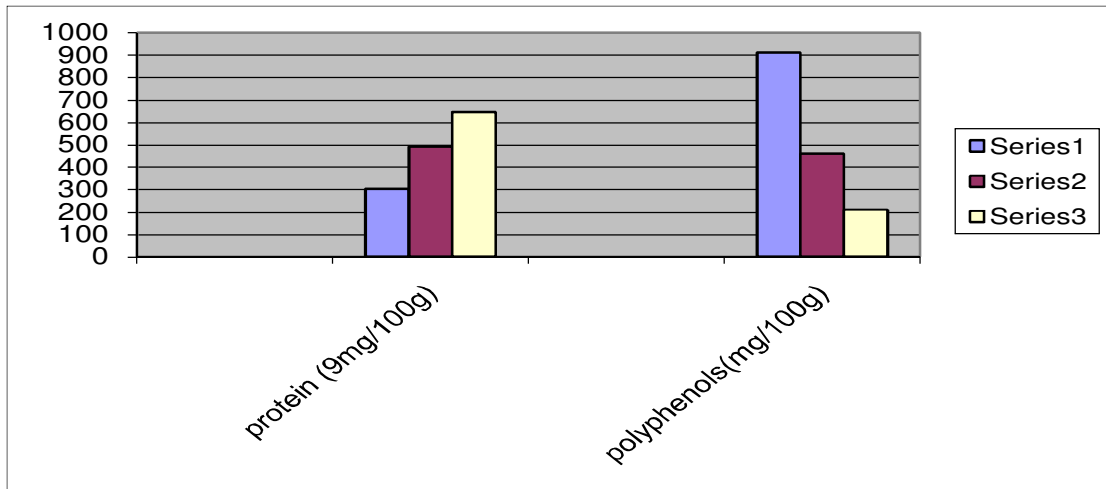


Fig. 4- Organic constituents (protein and polyphenols) in the different stages of regenerated shoots in sugarcane variety Co.7 (Series 1: IC, Series 2: IM and Series 3:- IR)

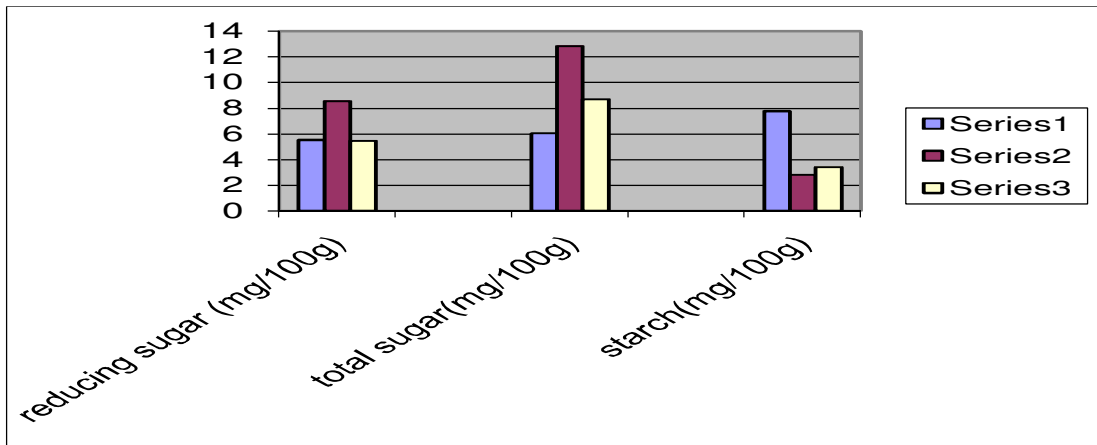


Fig. 5- Organic constituents (carbohydrates) in the different stages of regenerated shoots in sugarcane variety Co 740 (Series 1: IC, Series 2: IM and Series 3:- IR)