

BIOLOGICAL CONTROL OF FRUIT ROTS OF GRAPES CAUSED BY AUREOFUNGIN RESISTANT *ALTERNARIA TENIUS*

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Received: October 06, 2011; Accepted: November 30, 2011

Abstract- All the 16 fungal species from the phyllosphere, flower and the grape berries were tested for their antagonism against the resistant mutant of *Alternaria tenuis* (At-EMS-3) *Glaocladium roseum* and two bacterial isolates gave antagonistic reactions. Application of cultural filtrates from one of the bacterium was more useful for the management of pathogen on the berries when compared with dual culture or only the bacterial cells.

Key words- Biological control, Antagonism, At-EMS-3

Introduction- Grape (*Vitis vinifera* L.) is one of the very important fruit crop in India and Abroad. Grape plant become victim to many diseases in the life span but fruit rot is most important disease in the orchard and also in storage or transport. (Chahal and Mahli, 1969). Fruit rot of grapes caused by many fungi of these *Alternaria tenuis* fruit rot is most important one (Krishnaiah et al., 1983; Rao, 1993, 1994) fruit rot decreases market value of the grape fruit hence causing large economic loss to the farmer. The control of fruit rot of grapes has been almost exclusively based on the application of chemical fungicides, but use of fungicides is considered to be long term solution due to concern of expense risks, fungicide residue and other health and environmental hazards. Use of biocontrol agents has been shown to be ecofriendly and effective against many plant pathogens. It has been reported that *Fusarium sambucilum* (Kilkov et al., 1983), *Trichothecium roseum* (Dragoescu, 1986) prevent leaf and stem blight caused by *Alternaria solani*. The *Streptomyces* species and *Bacillus subtilis* (Mateascu et al., 2002; Sharma and Sharma, 2006; Sid et al., 2005) were also used to inhibit mycelial growth and spore germination of *Alternaria solani*. Hence attempt was made to find out the biological control of *Alternaria tenuis* causing fruit rot of grapes.

Materials And Methods- Isolates of different microbes were collected from fruits flower and phyllosphere of grape plant (*Vitis vinifera* L.) on Czapek-Dox agar medium poured in the petri plates. The resistant mutant of *Alternaria tenuis* (At-EMS-3) was inoculated on culture plates. Isolates of different fungi and bacteria were inoculated near *Alternaria tenuis* on same petri-plate and antagonistic reaction was observed.

Bacteria (*Bacillus* sp) were highly antagonistic to *Alternaria tenuis* so bacteria were used as Biological control for further studies.

In vitro effect of bacterial cultural filtrate on resistant mutant of *Alternaria tenuis* was observed by zone of inhibition. Antagonistic bacteria (*Bacillus* sp) were grown inside the nutrient broth medium in 100ml conical flask. Seven days after inoculation the cultural filtrate was obtained through cintered glass filter. Czapek-Dox agar medium poured in the sterilized petri plates and after solidification spore suspension of resistant mutant of *Alternaria tenuis* was spread on the surface of agar medium with the help of L shaped rod. A well was in the centre of agar medium with the help of cock borer (8mm diameter) 0.1 ml culture filtrate was poured in the well. After seven days of incubation zone of inhibition were observed and these zones were measured.

In vivo – Bacterial culture filtrates were obtained as above. Grape berries were washed 10 times with sterile distilled water. The berries treated with bacterial culture filtrate and inoculated with spore suspension of resistant mutant of *Alternaria tenuis*. These inoculated grapes were incubated at 26±3°C in moist Petri-plates and PDI was calculated. Without bacterial culture filtrate and bacterial cells were also used for treatment of grape berries and PDI was calculated.

Result And Discussion- A total of 16 fungal species isolated from various sources such as phyllosphere, flower and grape barriers were tested against *Alternaria tenuis* resistant mutant At- EMS – 3 on agar by spot inoculums plates. Various associative effects were seen. Results in Table-1 show that *Penicillium funiculosum*, *Aspergillus fumigates*, *Curvularia* sp., *Helminthosporeum* sp. and *Nigrospora* sp. did not show any harmful effect

against each other. *Alternaria tenuis* mutant overlapped on *Penicillium varians*, *Aspergillus flavus*, *Aspergillus niger* *Chaetomium sp.* and *Cunninghamella sp.* On the other hand *Rhizopus sp.*, *Cladosporium sp.*, *Trichoderma sp.* Overlapped on *Alternaria tenuis* mutant. *Gliocladium roseum* and two bacterial isolates indicated antagonistic zone towards *Alternaria tenuis* (Plate I). One of the bacterial isolates was used for biological control of *A. tenuis* mutants on agar plate and also on grape berries.

Effects of bacterial culture filtrate on resistant mutant of *A. tenuis* was observed by zone inhibition. Result in Table-2 and Plate II showed that inhibitory zone of bacterial culture filtrate was useful for the management of pathogen. The inhibitory zone of the two bacterial culture filtrates used were ranged from 11.5 to 13.5. Results in Table-3 showed that application of culture filtrate was more useful for management of this pathogen on the berries when compared with the dual culture and/or with the bacterial cells.

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Table 1: Antagonistic and associate relationship between *Alternaria tenuis* and other phyllosphere, flower and berries fungi of grapes.

| Sr no. | Name of the fungi | Reaction A | Reaction B | Reaction C | Reaction D |
|--------|--------------------------------|------------|------------|------------|------------|
| 1. | <i>Penicillium funiculosum</i> | + | - | - | - |
| 2. | <i>Penicillium varians</i> | - | + | - | - |
| 3. | <i>Aspergillus flavus</i> | - | + | - | - |
| 4. | <i>Aspergillus niger</i> | - | + | - | - |
| 5. | <i>Aspergillus fumigates</i> | + | - | - | - |
| 6. | <i>Curvularia sp.</i> | + | - | - | - |
| 7. | <i>Heminthosporium sp.</i> | + | - | - | - |
| 8. | <i>Nigrospora sp.</i> | + | - | - | - |
| 9. | <i>Chaetomium sp.</i> | - | + | - | - |
| 10. | <i>Cunninghamella sp.</i> | - | + | - | - |
| 11. | <i>Cladosporium sp.</i> | - | - | + | - |
| 12. | <i>Rhizopus sp.</i> | - | - | + | - |
| 13. | <i>Trichoderma sp.</i> | - | - | + | - |
| 14. | <i>Gliocladium roseum sp.</i> | - | - | - | + (2) |
| 15. | <i>Bacillus sp.</i> | - | - | - | + (3) |
| 16. | <i>Bacillus Sp.</i> | - | - | - | + (3) |

A = Both colonies grow side by side without harming each other
 B = *Alternaria tenuis* overlaps on other fungi.
 C = Other fungi overlaps on *Alternaria tenuis*.
 D = Other fungi antagonistic to *Alternaria tenuis*.
 *Figures in parentthesis show antagonistic zone in mm.

Table 2- Inhibitory action of bacterial culture filtrate against *Alternaria tenuis* (At- EMS-3) by cup plate method.

| Sr.No. | Bacterial culture filtrate | Inhibitory zone(mm) |
|--------|----------------------------|---------------------|
| 1. | <i>Bacillus</i> spp. | 11.5 |
| 2. | <i>Bacillus</i> spp. | 13.5 |

Table 3- Percentage control efficacy (PCE), *Bacillus* sp. Culture filtrate against Aureofungin resistant mutant of *Alternaria tenuis* on grape berries.

| Bacterial sources | PCE |
|-------------------|-------|
| Culture filtrate | 84.21 |
| Dual culture | 71.10 |
| Bacterial cells | 59.89 |
| S.E. | 5.73 |
| C.D. at 0.05 | 81.14 |
| C.D. at 0.01 | 85.04 |

**Plate- I-** Bacterium antagonistic to *A. tenuis* (At- EMS – 3)**Plate- II-** Inhibition zone of Bacterial culture filtrates to antagonistic to *A. tenuis* (At- EMS – 3) on Agar plates