



Research Article

STUDY OF ARSENIC UPTAKE BY THE RICE GENOTYPE IN RELATION TO NUTRIENT OXY-ANIONS

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Abstract: Rice is the main food crop is possibly cultivated on contaminated sites. A study was conducted to study of arsenic uptake by the rice genotype in relation to nutrient oxy-anions. Shoot arsenic concentrations under phosphate treatments were comparatively lower than under the corresponding concentration of nitrate and sulphate treatments under As stress condition. Root arsenic concentration decreased with increase in concentration of nitrate and phosphate in the nutrient. Increased level of nitrate through diluted arsenic concentration in root and shoot by promoting tissue growth it appeared to have little effect on uptake and translocation.

Keywords: Arsenic, Phosphate, Nitrate, Sulphate, Rice

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Introduction

Especially inorganic As species like arsenite and arsenate are highly carcinogenic posing a possible health risk to humans. Arsenic enters the human food chain mainly via drinking water or via food crops. Therefore rice has been the target cereal for investigating uptake and accumulation mechanisms in recent years [1,2]. Essential nutrient element like nitrogen and phosphorus have similarity in their atomic structure, all have five electron in outer orbit and group V element. Nitrogen, phosphorus and arsenic belong to the periods 2, 3, and 4 respectively. Sulphur is a group VI element of period 3. All these nutrient elements are taken up by plants as their oxy-anions, nitrogen as nitrate, phosphorus as phosphate and sulphur as sulphate. Interaction of these nutrient oxy-anions with arsenic oxy-anions is of course a worth study. So, the present experiment was carried out to study of arsenic uptake by the rice genotype in relation to nutrient oxy-anions [3].

Materials and Methods

This experiment was conducted in net house under the Directorate of Research, Bidhan Chandra Krishi Viswavidyalaya, Kalyani, Nadia. *Rasi* (IET-1444), a popular rice genotype and TN-1 female parent of *Rasi* were selected and grown hydroponically. Rice seeds (*Oryza sativa* L) cv. *Rasi* and TN-1 were surface sterilized with 0.1% (w/v) HgCl₂ for two minutes, washed repeatedly with glass distilled water, seeds were grown in pots for 35 days and after 35 days plants were transferred in Hoagland's solution. Seedlings with uniform growth and vigor were taken for experiment. The roots of such seedlings were properly washed initially with the tap water to remove the soil and other materials and finally with distilled water before transferring them in the culture solution.

Procedure of Planting

The roots of the seedlings were carefully inserted through the hole of the lid made up of thermocole and set on the top of the bucket. Staking was done to keep the seedling standing straight so that root part of the plant could reach the nutrient solution of the bucket. Every care was taken to avoid any physical damage of the seedling either at the root zones or in the aerial portion.

Preparation of culture solution

Basic culture solution was prepared by adding appropriate concentration of nutrients in Hoagland's solution (250 ml) (pH 5.5) containing (in mM): KNO₃ : 20; Ca(NO₃)₂, 2.0; MgSO₄, 0.7; and (in μM), Fe-EDTA, 0.50; ZnSO₄, 0.5; CuSO₄, 0.5; MnSO₄, 2.5; H₃BO₃, 5; Na₂MoO₄, 0.25; CuSO₄, 0.09. Arsenic was applied in the form of Sodium arsenate (Na₂HAsO₄·7H₂O, M. W. = 321.01) with 10 mg/l in which three levels of nutrient oxyanions in the form of MgSO₄, KNO₃, KH₂PO₄ (0.5 mM, 1.0 mM, 2.0 mM) were used. After 3 days plants were removed from hydroponics and placed in drier for arsenic analysis.

Analysis of the total arsenic in plant sample

Preparation and digestion of plant sample

After 3 days of treatment, leaves and root from the rice seedling sample were digested in an Erlenmeyer flask by a mixture of concentrated tri-acids, e.g., HNO₃, HClO₄ and H₂SO₄ in a proportion of 10:4:1 (v/v). The entire digestion process lasted 3-4 h. after complete digestion, the solution was diluted with double distilled water and filtered by Whatman No. 42 filter paper and transferred in to acid-washed plastic bottle; this solution was used for analyzing the arsenic and phosphorus content of the sample. Each treatment was performed in triplicate. The digest was diluted to 20 ml.

Determination of 'As' in plant sample

Two ml of the aliquot was taken in 10 ml plastic tube, 1 ml of concentrated HCl and 1 ml of mixed reagent [5% KI (w/v) +5% Ascorbic acid (w/v)] were added to it, kept for 45 minutes to ensure complete reaction and the volume was made up to 10 ml. The resultant solution was analyzed in a PerkinElmer Atomic Absorption Spectrophotometer with Flow Injection Analysis System (FIAS 400) @ λ_{max}193.7 nm where the carrier solution was 10% v/v HCl, the reducing agent (to ensure all As species be reduced to AsH₃ and to be measured against a calibration with standard As+3 solution) was 0.2% NaBH₄ in 0.05% NaOH (3)

Statistical analysis

The data of different parameters collected, were subjected to statistical analysis as per design(s) following the method described by 1 to find out the significance

between treatments used in the experiment. The experimental data for the characters were subjected to the variance analysis appropriate to a CRD design.

Shoot arsenic concentration (mg/kg) under different concentration of oxy-anion with As-V

The variations in shoot arsenic concentration due to genotypes, treatment and genotype treatment interaction were statistically significant such result indicates uptake varies with the genotypes, species and concentration of oxy-anion and also with interaction between genotype and treatment. Shoot As-V concentration of *Rasi* was higher than TN-1. Shoot As-V concentrations under contaminated treatments in combination with nutrient oxy-anions were lower than under only As-V treatment. It was comparatively lower under phosphate treatment than the other two oxy-anion treatments. Shoot arsenic concentrations under phosphate treatments were comparatively lower than under the corresponding concentration of nitrate and sulphate treatments. Shoot As concentrations were found to decrease with increase in concentration of each of the nutrient oxy-anions. The trend in shoot arsenic concentration of TN-1 was also similar to *Rasi* with only difference that it was lower than *Rasi* under every corresponding treatment.

Table-1 Shoot arsenic concentration under different concentration of oxy-anion with As-V

Treatments	Genotype		Mean
	<i>Rasi</i>	TN-1	
Control (Distilled water)	0.97	0.507	0.739
AS(V) 10 ppm	10.537	7.647	9.092
AS(V) 10mg/l+MgSO ₄ 0.5 mM	8.537	7.53	8.034
AS(V) 10mg/l+MgSO ₄ 1.0 mM	7.76	7.347	7.554
AS(V) 10mg/l+MgSO ₄ 2.0 mM	5.63	5.673	5.652
AS(V) 10mg/l+KNO ₃ 0.5 mM	8.373	5.65	7.012
AS(V) 10mg/l+KNO ₃ 1.0 mM	6.623	5.11	5.867
AS(V) 10mg/l+KNO ₃ 2.0 mM	6.14	4.4	5.27
AS(V) 10 mg/l+KH ₂ PO ₄ 0.5 mM	8.197	5.083	6.64
AS(V) 10 mg/l+KH ₂ PO ₄ 1.0 mM	6.177	5.04	5.609
AS(V) 10 mg/l+KH ₂ PO ₄ 2.0 mM	4.85	3.75	4.3
Mean	6.708	5.249	5.979
For comparison mean of	SEm (±)		CD (P = 0.05)
V	0.0523		0.1492
T	0.1228		0.3499
V × T	0.1737		0.4949

Table-2 Root Arsenic concentration under different concentration of oxy-anion with As-V

Treatments	Genotypes		Mean
	<i>Rasi</i>	TN-1	
Control (Distilled water)	11.493	5.873	8.683
AS(V) 10 ppm	59.867	52.927	56.397
AS(V) 10mg/l+MgSO ₄ 0.5 mM	67.78	70.07	70.425
AS(V) 10mg/l+MgSO ₄ 1.0 mM	68.027	70.367	69.197
AS(V) 10mg/l+MgSO ₄ 2.0 mM	68.327	70.413	69.37
AS(V) 10mg/l+KNO ₃ 0.5 mM	46.123	39.287	42.705
AS(V) 10mg/l+KNO ₃ 1.0 mM	40.363	38.847	39.605
AS(V) 10mg/l+KNO ₃ 2.0 mM	40.073	36.387	38.23
AS(V) 10 mg/l+KH ₂ PO ₄ 0.5 mM	50.897	40.85	45.873
AS(V) 10 mg/l+KH ₂ PO ₄ 1.0 mM	47.99	40.74	44.365
AS(V) 10 mg/l+KH ₂ PO ₄ 2.0 mM	40.193	39.517	39.855
Mean	49.194	46.207	47.7
For comparison mean of	SEm (±)		CD (P = 0.05)
V	0.5595		1.5944
T	1.3121		3.7393
V × T	1.8556		5.2882

Root Arsenic concentration (mg/kg) under different concentration of oxy-anion with AsV

The variations in root As concentration due to genotypes, treatment (nutrient oxy-anions of different concentration) and genotype treatment interaction were statistically significant. Such result indicates that uptake varies with the genotypes, species and concentration of oxy-anions and also with interaction between genotype and treatment. Average root As concentration of *Rasi* was higher than TN-1. Root As concentrations under As-V contaminated treatments in combination with nutrient oxy-anions except sulphate were lower than that under only As -V treatment. But it was higher than that under only As -V treatment in cases of

sulphate treatments in combination with As -V. It was lowest in cases of nitrate treatments. In *Rasi* root arsenic concentration decreased with increase in concentration of nitrate and phosphate in the nutrient medium whereas increased with increase in sulphate concentration in the nutrient medium. But in TN-1 change in concentration of the oxy-anion in the nutrient medium caused no significant difference in root arsenic concentration. Reduction in tissue arsenic concentration at higher level of phosphorous in the growing medium was also reported by Wang and Duan (2009) [4]. Arsenate was reported to compete directly with phosphate at uptake level [5,6]. Hence, high level of phosphorous in the growing media caused decline in tissue concentration of arsenic. Decline in translocation of arsenic under higher level of Sulphur is in agreement with the report made by Zhang *et al* (2010) who claimed that sulfur deprivation caused enhanced translocation of arsenic from root to shoot [7-9].

Conclusion

Shoot arsenic concentrations under phosphate treatments were comparatively lower than under the corresponding concentration of nitrate and sulphate treatments in arsenic contaminated condition. Increased Phosphate level caused greater decline in arsenic uptake as evidence by lower root and shoot arsenic concentration. Increased sulphate level caused little decline in shoot uptake but caused relatively greater decline in translocation. Increased level of nitrate though diluted arsenic concentration in root and shoot by promoting tissue growth it appeared to have little effect on uptake and translocation.

Application of research: This experiment is helpful in understanding and mitigating arsenic problems in field by application of nutrient oxyanions and efficiency of different oxyanions in solving arsenic problems.

Research Category: Plant Physiology

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References

- [1] Li R. Y., *et al.* (2009) *Plant Physiol.*, 150, 2071-2080.
- [2] Lombi E., *et al.* (2009) *New Phytol.*, 184, 193-201.
- [3] Schmidt D., Pitten F., Muller D. (2004) *Science of the total Environ.*, 226, 237-245.
- [4] Wang L. and Duan G. (2009) *J. of Environmental Sci.*, 21, 346-351.
- [5] Tu C., Ma L. Q. (2003) *Plant Soil*, 249, 373-382.
- [6] Wang J. R., Zhao F. J., Meharg A. A., Raab A., Feldman J. and McGrath S. P. (2002) *Plant Physiol.*, 130, 1552.
- [7] Panse D.G. and Sukhatme P.V. (1989) *Statistical methods for agricultural workers*. ICAR, New Delhi.
- [8] Zhao F.J., McGrath S.P. and Meharg A.A. (1910) *Annu. Rev. Plant Biol.*, 61, 535-559.
- [9] Zhang-Yi-Hu, Yang C. and Jian L. (2010) *Envtl. pollution*, 158, 409-415.