

Research Article

MOLECULAR DETECTION AND CHARACTERIZATION OF COAT PROTEIN GENE OF MUNGBEAN YELLOW MOSAIC VIRUS (MYMV) FROM KARNATAKA

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Abstract- Mungbean yellow mosaic disease is a major constraint in production of mungbean in Karnataka. An expected band size of approximately 1000 bp was amplified from MYMV-CP-F/MYMV-CP-R primers, designed specifically for MYMV. Mungbean yellow mosaic virus (MYMV) was characterized by sequencing the coat protein gene. The actual length of MYMV obtained was 889 bp including 115 bp pre-coat proteins at 5' end, 774 bp core coat protein and 257 deduced amino acids. The phylogenetic analysis of the coat protein sequence of yellow mosaic virus infecting mungbean was carried out together with the known begomoviruses sequences obtained from Genbank database. Cluster phylogram based comparison of the coding nucleotide sequences of MYMV-Hebbal-Bangalore isolate revealed that the MYMV-Hebbal, Bangalore isolate group with cluster II consisting of Mungbean yellow mosaic viruses. Analysis of nucleotide sequence of coat protein gene of yellow mosaic virus associated with mungbean showed maximum identity with 99% MYMV-Maharashtra: SB [AF314530.1], followed by 98.9% with MYMV-Namakka I: MoB [DQ865201.1] as well as MYMV-Tamil Nadu:MB [AJ132575.1] and 98.8% with MYMV-Madurai: SB isolates [AJ421642.1]. The nucleotide identity of MYMV-Hebbal isolate with other MYMV isolates ranged between 94.5-99%. MYMV-Hebbal, Bangalore isolate showed an identity of 79.3-81.0% with different MYMIV isolates. The amino acid sequence of mungbean yellow mosaic virus shared maximum identity of about 100% with MYMV-Maharashtra: SB [AF314530.1], MYMV-Tamil Nadu:MB [AJ132575.1], 99.6% with MYMV-Namakkal:MoB [DQ865201.1], 99.2% with MYMV-Madurai: SB [AJ21642.1]. The deduced amino acid identity of MYMV-Hebbal-Bangalore isolate with other MYMV isolates ranged between 96.4-100%. The aminoacid identity of MYMV-Hebbal-Bangalore isolate with other MYMIV isolates ranged between 96.4-100%. The aminoacid identity of MYMV-Hebbal-Bangalore is a Mungbean yellow mosaic virus (MYMV) but not Mungbean yellow mosaic locia virus (MYMIV) and is a variant of Mungbean yello

Keywords- Mungbean yellow mosaic virus (MYMV), Coat protein, Detection, Characterization, Phylogenetic analysis, Nucleotide sequence identity, Amino acid identity

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Introduction

Mungbean or Greengram [*Vigna radiata* (L.) Wilczek], is one of the food legumes grown in India. It is native to India and Central Asia. It consists of 23.6% of easily digestible protein and 51% carbohydrates. It is cultivated in China, Thailand, the Philippines, Vietnam, Indonesia, Myanmar, Bangladesh, India and in the hot and dry regions of Southern Europe and Southern United States [1]. It fixes biological nitrogen ranging from 30-74 kg/ha in the soil and also provides plant residues (15-20 quintals/ ha). Plants are also used for making green manures.

In India, mungbean is grown in an area of 33.87 lakh ha, with a production of 16.34 lakh tonnes and productivity of 483 kg/ha. The major mungbean producing states in India are Andhra Pradesh, Karnataka, Tamil Nadu, Maharashtra, Madhya Pradesh, Gujarat, Rajasthan and Orissa.

In Karnataka, the total area under mungbean is 2.93 lakh ha, giving a total production of 0.73 lakh tonnes, with an average productivity of 249 kg/ha. The major districts of Karnataka cultivating mungbean are Tumkur, Chickmagalur, Hassan, Shimoga, Mysore, Davanagere, Chitradurga, Mandya, Bidar, Kalaburagi and Yadgir[2].

The diseases affecting mungbean are cercospora leaf spot (*C. canescens, C. cruenta*), powdery mildew (*Erysiphe polygoni*), root disease complex (*Pythium* spp, *Rhizoctonia solani, Fusarium* spp.). Mungbean yellow mosaic virus disease

(MYMV) is the most damaging viral disease which was first reported from New Delhi in 1960. MYMV belongs to genus Begomovirus and family *Geminiviridae* [3]. It is transmitted mainly by whitefly, *Bemisia tabaci* (Genn.) in a circulative manner [4]. The virus has geminate particles (20 x 30 nm) with spherical, single stranded DNA fragment of 2.8 Kb [5]. The disease can even be transmitted by grafting but sap, seed or soil transmission is not seen [6]. The disease incidence ranged from 31.49 to 100 per cent in Southern Karnataka [7]. The virus inoculum is contributed by numerous alternate hosts of the virus and the whitefly vector, which help in rapid spread of the disease [8]. The characteristic symptoms of YMV on mungbean include irregular green and yellow patches in older leaves and complete yellowing of younger leaves. Fewer flowers and pods are produced by the infected plants. Pods of the affected plants often develop mottling, remain small and contain fewer and smaller seeds thus reducing the yield drastically [9, 10].

According to earlier reports, mungbean is affected by both Mungbean Yellow Mosaic Virus (MYMV) and Mungbean Yellow Mosaic India Virus (MYMIV). So, the investigations were carried out to prove that yellow mosaic virus infecting mungbean from Karnataka is an isolate of Mungbean yellow mosaic virus (MYMV) rather than Mungbean Yellow Mosaic India Virus (MYMIV). At present literature is available on the per cent disease incidence, epidemiology, virus-vector relationships, transmission, sources of resistance to MYMV and integrated

International Journal of Agriculture Sciences ISSN: 0975-3710&E-ISSN: 0975-9107, Volume 10, Issue 3, 2018 disease management practices [4, 6, 7, 11-24]. But the information at genomic level is not available for MYMV from Karnataka. The information on phylogenetic relationships of MYMV with other YMV isolates associated with other grain legumes are also not available. Hence, the present study has been taken up to characterize coat protein gene of MYMV infecting mungbean from Karnataka.

Material and Methods

Sample collection

Mungbean plants showing severe yellow mosaic and mottling symptoms were collected from field at the MRS, Hebbal, Bangalore, Karnataka during the 2012 [Plate-1]. Samples from healthy plants were collected as controls.



Plate-1 Mungbean plants showing typical symptoms of yellow mosaic virus

Nucleic acid extraction

The total genomic DNA was extracted from leaf tissues of healthy mungbean plants and YMV infected mungbean plants based on modified CTAB method [25]. One hundred and fifty milligrams of fresh YMV infected leaf tissues were ground with liquid nitrogen using sterile pestle and mortar. The whole ground sample was transferred into a fresh 1.5-ml eppendorf tube. 1500 µl of pre-warmed (65°C) DNA extraction buffer was added to ground sample taken in 1.5-ml eppendorf tube (added in situ just before DNA extraction). The whole crude sap was incubated for 30 min at 60° C in a water bath with occasional mixing. The supernatant (750 µl) was transferred into a fresh 1.5-ml effendorf tube and mixed with equal amount (750 µl) of Phenol: chloroform: isoamyl alcohol (25: 24:1) by vertexing. The samples were then centrifuged at 13,000 rpm for 10 min using micro centrifuge. The aqueous supernatant was collected in to a fresh 1.5-ml eppendorf tube. The DNA was precipitated by mixing with 300 µl of chilled isopropanol + 30 µl of 7.5 M Ammonium acetate by inversion. The tubes were centrifuged at 13,000 rpm for 10 min. The resulted pellet was washed with 70 per cent ethanol, dried in a vacuum drier for 10 min and re-suspended with 40 μ l of T₁₀E_{0.1} buffer (10 mM Tris-HCl of pH 8.0 and 0.1 mM EDTA of pH 8.0) and stored at -20° C. All the DNA extracts were further diluted from 1:10 to 1:40 in single distilled water (SDW) before using for PCR amplifications. The quality and quantity of DNA was assessed at 260 nm and 280 nm using UV spectrophotometer.

Primers used, PCR amplification and gel electrophoresis

In order to determine the nucleotide sequence of coat protein of mungbean yellow mosaic virus, specific primers available in the literature were tried to amplify coat protein region of yellow mosaic viruses of nearly 1000 bp. Primers specific to MYMV (MYMV-CP-F-ATG GG (T/G) TCC GTT GTA TGC TTG /MYMV-CP-R-GGC GTC ATT AGC ATA GGC AAT) were used for amplification of coat protein gene of mungbean yellow mosaic virus (MYMV). Primers were designed to get the complete coat protein gene of yellow mosaic viruses of legume hosts by taking 100 extra nucleotides on both the sides of the gene [26].

PCR was performed in Thermocycler (Eppendorf Master cycler gradient, Hamburg, Germany) programmed for one step of initial denaturation at 94° for 2 min and 35 cycles of denaturation at 94° C for 1 min, annealing at 55° C for 2 min for primers MYMV-CP-F/MYMV-CP-R and extension at 72° C for 3 min, followed by one step of final extension at 72° C for 10 min. PCR was conducted with Dream Taq Master mix (Fermentas) in total reaction mixture volume of 25 μ l that contained Dream *Taq* Master mix- 13 μ l; dH2O- 4 μ l; forward and reverse primers (20 pmole/ µl)- 2 µl each; DNA template (total nucleic acid-100ng/µl)- 4 µl, and PCR products were subjected to electrophoresis in 1 % agarose at 50 V for 45 minutes in Electrophoresis system - SCOTLAB (Anachem Ltd.) in Tris-acetate-EDTA buffer containing ethidium bromide @ 0.1 %. The gel was observed under Gel Documentation System (IMAGO Compact Imaging System, B & L Systems, Isogen Life science, The Netherlands).

Cloning and sequencing of coat protein gene of YMV infecting mungbean

The PCR products were purified from agarose gel using Qiagen Gel Extraction kit (Qiagen, Hilder, Germany). All amplicons were cloned into the plasmid vector pTZ57R/T using InsTAclone™ PCR Cloning Kit following the manufacturer's instructions. Transformed colonies were screened and selected on LB agar medium amended with ampicillin, X-gal and IPTG. Isolated plasmids from transformed positive clones were confirmed for the presence of insert using the respective CP specific primers. The resultant positive clones were fully sequenced in both directions using universal M13 forward and reverse primers. Full length sequence of coat protein of YMV was obtained by aligning of forward and reverse reaction sequences.

Phylogenetic analysis, nucleotide sequence and amino acid sequence comparison of coat protein gene of yellow mosaic virus of mungbean with other Gemini viruses

Pair wise and multiple sequence alignment of the full length of coat protein sequence of various YMV was done using MEGA 5.1 multiple alignment tool. The phylogenetic neighbor-joining trees and evolutionary analysis were conducted using MEGA 5.1 software package based on coat protein gene sequences of MYMV with 21 other Gemini virus sequences downloaded from NCBI Genbank [Table-1] [27]. Robustness of trees was determined by bootstrap sampling of multiple sequence alignment with 1000 replications. Comparison of the nucleotide and amino acid sequences of YMV was analysed by using sequence identity matrix tool of Bio-Edit software (Version 7.9.1).

Results and Discussion

The total nucleic acid was extracted from the leaves of mungbean with characteristic yellow mosaic symptoms using modified CTAB protocol. Primers used for the detection of MYMV were MYMV-CP-F/MYMV-CP-R corresponding to coat protein gene. The total DNA isolated from infected samples was used for PCR amplification. Polymerase chain reaction was employed to establish association of begomovirus through amplification of Gemini virus specific PCR product. Various dilutions 1:10 to 1:40 were prepared and subjected to PCR. The PCR results indicated that the virus could be detected from 1:20 to 1:40 dilutions. Virus specific DNA fragments of approximately 1000 bp were obtained from DNA of mungbean yellow mosaic infected samples, whereas no PCR product was obtained from DNA extracted from healthy samples and water control. PCR was performed in Thermocycler programmed for one step of initial denaturation at 94° C for 2 min and 35 cycles of denaturation at 94°C for 1 min, annealing at 55° C for 2 min and extension at 72° C for 3 min, followed by one step of final extension at 72° C for 10 min. An annealing temperature of 55° C for 2 min was found suitable for amplification of coat protein gene of mungbean yellow mosaic virus. PCR products were subjected to electrophoresis in 1 % agarose. The gel was observed under Gel Documentation System.

Detection of yellow mosaic virus infecting mungbean was employed by PCR assays. Primers specific to MYMV (MYMV-CP-F/MYMV-CP-R) corresponding to coat protein gene were used. An expected band size of approximately ~1000 bp was amplified from MYMV-CP-F/MYMV-CP-R primers, designed specifically for MYMV but not from healthy plants [Plate-2]. Different authors have used different set of primers which were designed to amplify coat protein region of different legume yellow mosaic viruses [26, 28-32]. The amplicons obtained were electrophoresed through 1% agarose gel in1X TAE and bands were visualized under UV light after staining with ethidium bromide (0.05 μ g/ml). The bands were later excised from the gel and eluted through Qiagen gel extraction kit and cloned into plasmid vector pTZ57R/T using the PCR cloning kit following the manufacturer's instructions. Plasmid purification was carried out

International Journal of Agriculture Sciences ISSN: 0975-3710&E-ISSN: 0975-9107, Volume 10, Issue 3, 2018 using a Qiagen plasmid miniprep kit. The insert was sequenced, assembled and its total length was found to be 889 bp. The actual length of Mungbean yellow mosaic virus isolate of Bangalore was 889 bp including 115 bp pre-coat protein at 5' end and 774 bp core coat protein. The length of deduced amino acid was 257.



Plate-2 Amplification of coat protein gene of YMV infecting mungbean using MYMV-CP-F/MYMV-CP-R primer pair

Lane:

M-1Kb Marker (NEB 1 kb DNA ladder)

Lane 1 - Healthy mungbean plant DNA

Lane 2, 3, 4 - Specific PCR product of 1000 bp from MYMV infected sample Lane 5 - Water control

The phylogenetic analysis of the coat protein sequence of yellow mosaic virus infecting mungbean was carried out together with the known begomoviruses sequences obtained from GenBank database. Gen Bank accession numbers of different begomoviruses used for MYMV-Bangalore isolate sequence comparison and phylogenetic analysis were presented in [Table-1].

Phylogentic tree based on alignment of coding nucleotide sequences of majority of Mungbean yellow mosaic viruses and Mungbean yellow mosaic India viruses showed two clusters with Mungbean yellow mosaic India virus(MYMIV) isolates falling in cluster I and Mungbean yellow mosaic virus(MYMV) isolates in cluster II [Fig-1]. Cluster phylogram based comparison of the coding nucleotide sequences of MYMV-Hebbal-Bangalore isolate revealed that the MYMV-Hebbal, Bangalore isolate group with cluster II. The present isolate clustered with MYMV-Nammakal, MYMV-Madurai, MYMV-Maharashtra and MYMV-Tamil Nadu isolates infecting moth bean, soybean, soybean and mungbean, respectively. The cluster phylogram based on multiple alignment of the nucleotide sequence of the CP gene of 10 isolates of MYMIV indicated that all the 10 isolates belonged to MYMIV as they formed cluster with other known isolates of MYMIV [32].

Table-1 List of geminiviruses used for comparision of coat protein gene sequences, their origin, host species and NCBI accession numbers							
SI. No.	Virus species	Abbreviation	Geographical origin	Host species	Accession number		
1.	Mungbean yellow mosaic virus	MYMV-Haryana:MB	Haryana	Mungbean(MB)	AY271896.1		
2.	Mungbean yellow mosaic virus	MYMV-Namakkal:MoB	Namakkal	Mothbean(MoB)	DQ865201.1		
3.	Mungbean yellow mosaic virus	MYMV:Combodia:MB	Combodia	Mungbean(MB)	AY271892.1		
4.	Mungbean yellow mosaic virus	MYMV-Madurai:SB	Madurai	Soybean (SB)	AJ421642.1		
5.	Mungbean yellow mosaic virus	MYMV-Pakistan:SB	Pakistan	Soybean(SB)	AY269991.1		
6.	Mungbean yellow mosaic virus	MYMV-Maharashtra:SB	Maharashtra	Soybean(SB)	AF314530.1		
7.	Mungbean yellow mosaic virus	MYMV-Thailand:MB	Thailand	Mungbean(MB)	AB017341.1		
8.	Mungbean yellow mosaic virus	MYMV-Tamil Nadu:MB	Tamil Nadu	Mungbean(MB)	AJ132575.1		
9.	Mungbean yellow mosaic India virus	MYMIV-Indonesia:SB	Indonesia	Soybean(SB)	JN368438.1		
10.	Mungbean yellow mosaic India virus	MYMIV-Akola:MB	Akola	Mungbean(MB)	AY271893.1		
11.	Mungbean yellow mosaic India virus	MYMIV-India:SB	India	Soybean(SB)	AY049772.1		
12.	Mungbean yellow mosaic India virus	MYMIV-Indonesia:YLB	Indonesia	Yard long bean (YLB)	JN368437.1		
13.	Mungbean yellow mosaic India virus	MYMIV-Pakistan:BG	Pakistan	Blackgram(BG)	FM208845.1		
14.	Mungbean yellow mosaic India virus	MYMIV-Pakistan:MB	Pakistan	Mungbean(MB)	AY269992.1		
15.	Mungbean yellow mosaic India virus	MYMIV-Indonesia:YLB	Indonesia	Yard long bean (YLB)	JN368434.1		
16.	Mungbean yellow mosaic India virus	MYMIV-Indonesia:YLB	Indonesia	Yard long bean (YLB)	JN368432.1		
17.	Mungbean yellow mosaic India virus	MYMIV-Nepal:MB	Nepal	Mungbean(MB)	AY271895.1		
18.	Mungbean yellow mosaic India virus	MYMIV-Varanasi:Do	Varanasi	Fieldbean(Do)	AY547317.1		
19.	Mungbean yellow mosaic India virus	MYMIV-Bangladesh:MB	Bangladesh	Mungbean(MB)	AF314145.1		
20.	Mungbean yellow mosaic India virus	MYMIV-Jabalpur:SB	Jabalpur	Soybean(SB)	AJ416349.1		
21.	Mungbean yellow mosaic India virus	MYMIV-Palampur:FB	Palampur	Frenchbean (FB)	FN794200.1		



Fig-1 Phylogenetic tree obtained from comparision of complete nucleotide sequence of coat protein gene of MYMV with other geminiviruses from database. The dendrograms are calculated using neighbor-joining algorithm of MEGA 5.1 version. Numbers at nodes indicate percentage bootstrap confidence scores (1,000 replications).

Analysis of nucleotide sequence of coat protein gene of yellow mosaic virus associated with mungbean showed maximum identity of 99 per cent with MYMV-Maharashtra:SB [AF314530.1], followed by 98.9 per cent with MYMV-Namakkal: MoB [DQ865201.1] as well as MYMV-Tamil Nadu:MB [AJ132575.1] and 98.8 per cent with MYMV-Madurai:SB [AJ421642.1] isolates. The identity of MYMV-Hebbal isolate with other MYMV isolates ranged between 94.5-99 percent. MYMV-Hebbal, Bangalore isolate showed an identity of 79.3-81.0 per cent with different MYMIV isolates [Table-2]. Analysis of CP gene sequences of isolates from mungbean and urdbean genotypes revealed these isolates had 95-99 per cent similarity at nucleotide level with known MYMIV isolates [32].

The amino acid sequence of mungbean yellow mosaic virus shared maximum identity of about 100 per cent with MYMV-Maharashtra:SB [AF314530.1], MYMV-Tamil Nadu:MB [AJ132575.1], 99.6 per cent with MYMV-Namakkal:MoB [DQ865201.1] and 99.2 per cent with MYMV-Madurai:SB [AJ421642.1]. The deduced amino acid identity of MYMV-Hebbal-Bangalore isolate with other MYMV isolates ranged between 96.4-100 per cent. The amino acid identity of MYMV-Hebbal-Bangalore isolate with other MYMV-Hebbal-Bangalore isolate with other MYMV-Hebbal-Bangalore isolate with other MYMV-Hebbal-Bangalore isolate with other MYMV isolates ranged from 84.0-85.9 per cent [Table-2]. Analysis of CP gene sequences of isolates from mungbean and urdbean genotypes revealed that these isolates had 96-100 per cent similarity at

International Journal of Agriculture Sciences ISSN: 0975-3710&E-ISSN: 0975-9107, Volume 10, Issue 3, 2018 amino acid level with known MYMIV isolates [32]. The above results revealed that yellow mosaic virus infecting mungbean from Hebbal, Bangalore is a Mungbean yellow mosaic virus but not Mungbean yellow mosaic India virus and is a variant of Mungbean yellow mosaic virus. The results obtained are in agreement with earlier investigations carried out by some other authors [26, 28, 30-33].

Table-2 Nucleotide and amino acid sequence identities of coat protein gene of vellow mosaic virus infecting greengram with other geminiviruses

SI. No.	Accession number	Sequences	Nucleotide sequence identity	Amino acid sequence identity
1.	AY271896.1	MYMV-Haryana:MB	94.5	96.4
2.	DQ865201.1	MYMV-Namakkal:MoB	98.9	99.6
3.	AY271892.1	MYMV:Combodia:MB	97.1	98.0
4.	AJ421642.1	MYMV-Madurai:SB	98.8	99.2
5.	AY269991.1	MYMV-Pakistan:SB	95.0	97.2
6.	AF314530.1	MYMV-Maharashtra:SB	99.0	100
7.	AB017341.1	MYMV-Thailand:MB	97.4	98.8
8.	AJ132575.1	MYMV-Tamilnadu:MB	98.9	100
9.	JN368438.1	MYMIV-Indonesia:SB	79.9	84.0
10.	AY271893.1	MYMIV-Akola:MB	79.3	84.4
11.	AY049772.1	MYMIV-India:SB	81.0	85.6
12.	JN368437.1	MYMIV-Indonesia:YLB	80.1	85.2
13.	FM208845.1	MYMIV-Pakistan:BG	80.1	85.6
14.	AY269992.1	MYMIV-Pakistan:MB	80.2	85.9
15.	JN368434.1	MYMIV-Indonesia:YLB	79.9	84.4
16.	JN368432.1	MYMIV-Indonesia:YLB	79.9	84.8
17.	AY271895.1	MYMIV-Nepal:MB	79.5	85.2
18.	AY547317.1	MYMIV-Varanasi:Do	79.5	85.6
19.	AF314145.1	MYMIV-Bangladesh:MB	80.1	84.0
20.	AJ416349.1	MYMIV-Jabalpur:SB	79.8	85.9
21.	FN794200.1	MYMIV-Palampur:FB	79.8	85.9

Conclusions

Mungbean is affected by both Mungbean Yellow Mosaic Virus (MYMV) and Mungbean Yellow Mosaic India Virus (MYMIV) according to earlier reports whereas our study reports the infection of MYMV on Mungbean from Karnataka. The CP gene is the most highly conserved gene in the family *Geminiviridae*. The CP sequence effectively predicts discrete strains, species and taxonomic lineage of begomoviruses. The International Committee on Taxonomy of Viruses (ICTV) accepts the classification of begomoviruses based on CP gene sequences. Member of the genus begomovirus are known to form clusters according to geographical origin. The results of the phylogenetic analysis, nucleotide sequence comparison and amino acid sequence comparison revealed that coat protein gene of yellow mosaic virus infecting green gram (MYMV-Hebbal-Bangalore) is a Mungbean yellow mosaic virus (MYMV) but not Mungbean yellow mosaic India (MYMIV) virus and it is a variant of mungbean yellow mosaic virus since it showed 94.5-99 per cent identity at nucleotide level with other MYMV isolates.

Application of research

The results of the present study help in devising effective management strategies against MYMV by developing transgenics based on coat protein or gene silencing approaches.

Research Category: Agriculture/Plant Pathology/Plant Virology

Abbreviations

MYMV-Mungbean yellow mosaic virus MYMIV-Mungbean yellow mosaic India virus CP-Coat protein PCR-Polymerase chain reaction

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Conflict of Interest: None declared

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References

- Nair R. M., Yang R. Y., Easdown W. J., Thavaraiah D., Thavaraiah P. and Hughes J. d.'A. (2013) *Journal of the Science of Food and Agriculture*, 93, 1805-1813.
- [2] Anonymous (2012) Selected state wise Area, Production and Productivity of Moong (*Kharif* and *Rabi*) in India, Ministry of Agriculture and Farmers Welfare. Govt. of India.
- [3] Bos L. (1999) Plant Viruses: Unique and Intriguing Pathogens: A Text Book of Plant Virology, Backhuys Publishers, The Nether-lands, 305-306.
- [4] Nair N.G. and Nene Y.L. (1973) Indian Journal of Farm Science, 1, 109-110.
- [5] Roger Hull (2004) *Mathew's Plant Virology*, 4th ed., Elsevier Pub-lishers, India, 180-182.
- [6] Nariani T.K. (1960) Indian Phytopathology, 13, 24-29.
- [7] Manjunath B., Jayaram N., Muniyappa V. and Prameela H. A. (2013) Legume Res., 36(1), 62-66.
- [8] Malathi V. G. and John P. (2008) Gemini viruses infecting legumes. In: G. P. Rao, P. L. Kumar, and R. J. Holguin–Pen^a (Eds.), Vegetable and pulse crops: Vol. 3. Characterization, diagnosis and management of plant viruses (pp. 97–123). USA: Studium Press LLC.
- [9] Nene Y.L. (1973) Plant Disease Reporter, 57, 463-467.
- [10] Dhingra K.L. and Chenulu, V.V. (1985) Indian Phytopathology, 38, 248-251.
- [11] Capoor S.P. and Varma P.M., (1950a) Current Science, 19: 248-249.
- [12] Chenulu V.V. and Verma A. (1988) Virus and virus like diseases of pulse crops commonly grown in India. In: *Pulse Crops*, Eds. Baldev, B., Ramanujam, S. and Jain, H.K., Oxford and IBH Publishing Co., New Delhi, pp.338-370.
- [13] Chenulu V.V., Venkateswarlu V. and Rangaraju R. (1979) Indian Phytopathology, 32, 230-235.
- [14] Deepa H., Govindappa M.R., Sunil Kulakarni Kenganal M. and Biradar S.A. (2017) Int. J. Curr. Microbiol. App. Sci., 6(10), 678-684.
- [15] Ahmed M. and Harwood R.F. (1973) Plant Disease Reporter, 57: 800-802.
- [16] Jayappa, Ramappa H. K., Jabbar Sab and Devamani B. D. (2017) Int. J. Pure App Biosci., 5 (3), 238-244.
- [17] Murugesan S. and Chelliah S. (1977) Madras Agricultural Journal, 64(2), 128-130.
- [18] Nath P.D. (1994) Annals of Agricultural Research, 15(2), 174-177.
- [19] Pathak A.K. and Jhamaria S.L. (2004) Journal of Mycology and Plant Pathology, 34(1), 64-65.
- [20] Peerajade D.A., Ravikumar R.L. and Rao M.S.L. (2004) Indian Journal of Pulses Research, 17(2), 190-191.
- [21] Rathi Y.P.S. and Nene Y.L. (1974a) Indian Phytopathology, 27, 459-462.
- [22] Rathi Y.P.S. and Nene Y.L., (1974b) Acta Botanica Indica, 2, 74-76.
- [23] Rathi Y.P.S. and Nene Y.L. (1976) Pantnagar Journal of Research, 1, 107-111.
- [24] Shaik Abdul Salam (2005) Studies on mungbean yellow mosaic virus disease on greengram. *M.Sc. (Agri.) Thesis*, Univ. Agric. Sci, Dharwad, 75.

- [25] Rouhibakhsh A., Priya J., Periasamy A., Haq Q. M. I. and Malathi V. G. (2008) *J. Virological Methods*, 147, 37-42.
- [26] Naimuddin and Mohd. Akram (2010) J. Food Legumes, 23, 191-195.
- [27] Tamura K., Dudley J., Nei M. and Kumar S. (2007) Mol. Biol. Evol., 24, 1596-1599.
- [28] Obaiah S. (2011) Molecular detection and characterization of yellow mosaic virus infecting blackgram in Andhra Pradesh. *M. Sc. (Agri.) Thesis*, Acharya N.G. Ranga Agri. Univ., Hyderabad, 99pp.
- [29] Naimuddin, Mohd. Akram, Aditya Pratap, Brijesh Kumar Chaubey and Joseph K. John (2011) *J. Food Legumes*, 24, 14-17.
- [30] Kamaal Naimuddin, Mohammad Akram and Gupta Sanjeev (2011) *Phytopathol. Mediterr.*, 50, 94-100.
- [31] Mohammad Nurul Islam, Sonia Khan Sony and Rita Sarah Borna (2012) *Pl. Tissue Cult. Biotech.*, 22, 73-81.
- [32] Naimuddin and Akram M. (2012) J. Food Legume, 25, 286-290.
- [33] Sachan Mansi, Mishra Minakshi, Naimuddin and Akram Mohd. (2010) *Trends Biosci.*, 3, 166-168.