



Research Article

PRIMER DESIGNING AND VALIDATION OF STARCH PHOSPHORYLASE GENE FROM *VIGNA SINENSIS* VARIETY PUSA KOMAL

CHAUHAN RANJEETA AND MAHATMAN KRITYANAND KUMAR*

School of Biotechnology, Devi Ahilya University, Khandwa Road, Indore, MP, 452017, India

*Corresponding Author: Email-kriyabt@gmail.com

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Abstract- Primer designing for Starch Phosphorylase Gene is done using Primer 3 software, with wet lab validation work has been carried out by PCR operation with agarose gel electrophoresis. Similar sequences for Starch Phosphorylase (*Vigna sinensis*) were retrieved from NCBI's RefSeq database through Entrez, on multiple sequence alignment, conserved clusters were identified. The potential primers were 18-24 nucleotides selected in this region by all combination. The potential primer was analyzed using Primer Biosoft's Net primer tool. The forward primer having 5' end and backward primer having 3' end and 50 to 60% GC content, and 54 to 58^oC T_m and without secondary structure were finalized. Specificity of the Primer was validated by carrying out the local alignment against the NCBI nr database through BLAST. All the alignment showed significant alignment to starch validating the specificity of the Primer.

Keywords- Starch phosphorylase, Primer designing, *Vigna Sinensis*.

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Introduction

Computer technology helps in the management of biological information; it offers computational, molecular biologists and another research scientist an open and extensible environment in which to explore ideas, prototype, and new algorithms. Bioinformatics Toolbox software provides access to genomics and proteomic data formats, analysis technique and specialized visualization for genomic, proteomic sequence and microarray analysis. It also includes collection, organization, storage and retrieval of biological information from the databases selection of oligonucleotide primers for polymerase chain reaction (PCR) oligo-hybridization and DNA sequencing. Proper primer design is actually one of the most important factors in successful DNA sequencing. For maximal specificity and efficiency of polymerase chain reaction appropriate primer concentration and optimal primer sequence is necessary. A poorly designed primer having little or no product due to nonspecific amplification and primer dimer formation having competitive enough to suppress product formation. There are several online tools available that are devoted to serving molecular biologist for effective PCR primer design [1].

Starch phosphorylase (EC. 2.4.1.1); alpha-1, 4-glucan: orthophosphate, alpha-glucosyl transferase) catalyzes the reversible conversion of starch into glucose-1-phosphate in the presence of orthophosphate. Normally starch consists of two components amylose and amylopectin; both are the polymer of glucose molecules. Amylose is a linear alpha-1, 4 D-glucan polymers and amylopectin is branched alpha-1, 6 D-glucan polymers. Starch phosphorylase is the first enzyme in the degradation of starch in plants. Starch phosphorylase is widely distributed in the plant kingdom [2-4] and has been demonstrated in several plant parts. Multiple forms of the enzyme have also been suggested as a biochemical response to the heterogeneity of starch molecules and starch granule structure [5]. Multiplicity may occur at the post-transcriptional level due to presence of different genes or at the post-transcriptional level due to differences in the splicing of mRNA. It can be used for the production of glucose-1-phosphate a cytostatic compound used in cardio-

therapy [6-7]. Starch phosphorylase used to estimate the amount of inorganic phosphate in serum under the pathological conditions and amount of inorganic phosphate pollution in the environment detected. After considering the importance of starch phosphorylase enzyme in the present study, we have designed primers for starch phosphorylase genes validation of the designed primer is done through PCR.

Materials and Methods

Seeds of *Vigna sinensis* variety Pusa Komal were collected from a farm. Seeds were germinated in plant growth chamber at 28°C and 60% relative humidity. After 3 days, the germinated seeds were used for further work.

Web-based resources for primer design: There are numerous Web-based resources for PCR primer design. Though most are freely available, they are of variable quality and not well maintained. Therefore, web-based resources often result in missing links and Web sites that have been useful previously may not be functional at a later date. So many criteria such as primer length, GC contents, T_m, dimer formation false priming, hair pin loop should be established in design of primers [1].

Software in primer design:

The use of software in the biological application has given a new dimension to the field of bioinformatics. Many different programs for the design of primers are now available. Free software is available on the internet and many universities have established servers where a user can log on and perform free analyses of proteins and nucleic acid sequences. There is a number of the stand-alone program as well as complex integrated networked version of the commercial software available. These software packages may be used for secondary structure predictions, DNA and protein analysis, primer designing, molecular modeling,

Specificity of the primers was validated by carrying out the local alignment against the NCBI's nr database through BLAST [Fig-3]. The BLAST search was carried against the non-redundant (nr) database. The designed forward and backward primer can be used for PCR amplification of starch Phosphorylase Gene for *Vigna sinensis*.

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> amb|X52385.1| Potato mRNA for starch phosphorylase
Length=4942
Score = 42.1 bits (21), Expect = 0.021
Identities = 21/21 (100%), Gaps = 0/21 (0%)
Strand=Plus/Plus
Query 1 CCTGAGGCACTGGAGAAATGG 21
      |||
Sbjct...3190 CCTGAGGCACTGGAGAAATGG 3210

> db|D00520.1|POTGP Solanum tuberosum mRNA for alpha-glucan phosphorylase
precursor,
Complete cds
Length=3101

Score = 42.1 bits (21), Expect = 0.021
Identities = 21/21 (100%), Gaps = 0/21 (0%)
Strand=Plus/Plus
Query 1 CCTGAGGCACTGGAGAAATGG 21
      |||
Sbjct 1331 CCTGAGGCACTGGAGAAATGG 1351

> gb|L25626.1|IPBSTAPHOA Ipomoea batatas starch phosphorylase gene,
complete cds
Length=6594

Score = 42.1 bits (21), Expect = 0.021
Identities = 21/21 (100%), Gaps = 0/21 (0%)
Strand=Plus/Plus
Query 1 CCTGAGGCACTGGAGAAATGG 21
      |||
Sbjct 3389 CCTGAGGCACTGGAGAAATGG 3409

> gb|M64362.1|IPBSPP Sweet potato starch phosphorylase mRNA, complete cds
Length=3292

Score = 42.1 bits (21), Expect = 0.021
Identities = 21/21 (100%), Gaps = 0/21 (0%)
Strand=Plus/Plus
Query 1 CCTGAGGCACTGGAGAAATGG 21
      |||
Sbjct...1352 CCTGAGGCACTGGAGAAATGG 1372
```

Fig-3 BLAST results for designed primers. Primer showed alignment to starch phosphorylase gene.

Table-1 Various properties of four forward and four backward primers analyzed using the net primer

Name of the primer	Sequence	Hairpin	Dimer	Tm °C	GC%
F1	CCTGAGGCACTGGAGAAATGG	0	0	61.88	57.14
F2	TGAGGCACTGGAGAAATG	0	0	51.26	50
F3	GAGGCACTGGAGAAATGG	0	0	52.73	55.56
F4	GATGCTGCTCTAGGAAATGG	0	0	61.3	50
R1	ACGACATTTACCTCAACGGC	0	0	57.59	50
R2	CGACATTTACCTCAACGG	0	0	51.47	50
R3	GACATTTACCTCAACGGC	0	0	50.42	50
R4	TACCCTGGAATTGGCACATC	0	0	64.5	50

DNA Isolation and PCR amplification: DNA was extracted by CTAB miniprep methods. 7 µl of DNA sample from stock was diluted with 35 µl of double distilled water, 10 µl of this diluted DNA was loaded on 0.8 percent agarose gel and electrophoresis was done for about 2 hours at the constant current of 50 volts. The gel was observed on UV-transilluminator and picture was recorded on a gel documentation system. 5 µl of DNA from stock was diluted up to 20 times using 95 µl of double distilled water (nanopore). The diluted DNA was subsequently used for PCR amplification. PCR reactions were performed in the total volume of 25 µl reaction volume containing PCR buffer 1X with 1.5 mM MgCl₂, 0.25mM dNTPs, 0.100 µM of each primer, 3U Taq polymerase and 50 ng of genomic DNA as a template. After an initial denaturing step of 2 min at 94°C, 35 cycles were performed for 30 sec at 94°C, 51°C for 1 minute and 72°C for 1 minutes, with a final extension at 72°C for 5 min.

PCR amplification was generated with a set of 4 primers. The primer used for this purpose is listed in [Table-1]. PCR amplified product was (mixed with 6x loading dye and loaded on 3 per cent Agarose) prepared in 1x TBE buffer along with 100bp plus ladder. Electrophoresis was done for 2 hours at 120 volts. Gels were visualized on UV transilluminator and photographed by using SynGene gel documentation system.



Fig-4 PCR amplification using all four primers sequence Ladder, P1 (4 sample) P2 (4 sample) P3 (4 sample) P4 (4 sample), Ladder

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Conclusion: These research finding may be useful in future for cloning and expression of genes.

Author Contributions: Both author equally contributed

Abbreviations: PCR: Polymerase Chain Reaction, BLAST: Basic Local Alignment Search Tool, NCBI: National Centre for Biological Information

Ethical approval: This article does not contain any studies with human participants or animals performed by any of the authors.

Conflict of Interest: None declared

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